Chapter 6: Antigen-Antibody Interactions

I. Strength of Ag-Ab interactions A. Antibody Affinity

> - strength of total noncovalent interactions between single Ag-binding site on an Ab and a single epitope is the affinity of the Ab for that epitope

- low affinity Ab: bind Ag weakly and dissociate readily

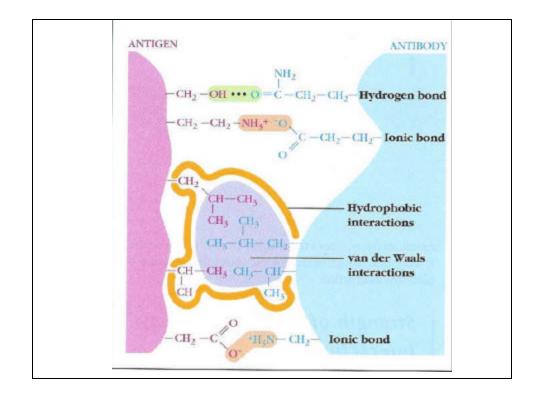
- high affinity Ab: bind Ag tightly and remain bound longer

- Ka (measure of Ab affinity) can be calculated from the ratio of the molar concentration of bound Ag-Ab complex to the molar concentrations of unbound Ag and Ab at equilibrium

- Ka = [Ag-Ab] / [Ab][Ag]

- rate at which bound Ag leaves the Ab's binding site helps determine the Ab's affinity for an Ag

- noncovalent forces involved in Ag-Ab interaction: hydrogen bonds, ionic bonds, hydrophobic interactions, and van der Waals forces



B. Antibody Avidity

strength of multiple interactions between a multivalent Ab and Ag
avidity of an Ab is a better measure of its binding capacity within biological systems than the affinity of its individual binding sites

high avidity can compensate for low afinity

II. Cross-Reactivity

Ab elicitied by one Ag can cross-react with an unrelated Ag
cross-reactivity occurs if 2 different Ag's share

an identical epitope or if Ab's specific for one epitope also bind to an unrelated epitope possessing similar chemical properties

- example of cross-reactivity:
 - ABO blood-group antigens

- subtle differences in the terminal sugar residues distinguish the A and B bloodgroup antigens

- cross-reactivity is the basis for the presence of these blood-group antibodies, which are induced in an individual not by exposure to red blood cell antigens but by exposure to cross-reacting microbial antigens present on common intestinal bacteria

TABLE 6-2 ABO BLOOD TYPES		
Blood type	Antigens on RBCs	Serum antibodies
-	А	Anti-B
A		
A B	В	Anti-A
	B A and B	Anti-A Neither

III. Precipitation Rections

- antibodies that aggregate soluble antigens are called precipitins

- formation of an Ag-Ab lattice depends on the valency of both the Ab and Ag:

- the Ag must be bivalent; a precipitate will not form with monovalent Fab fragments

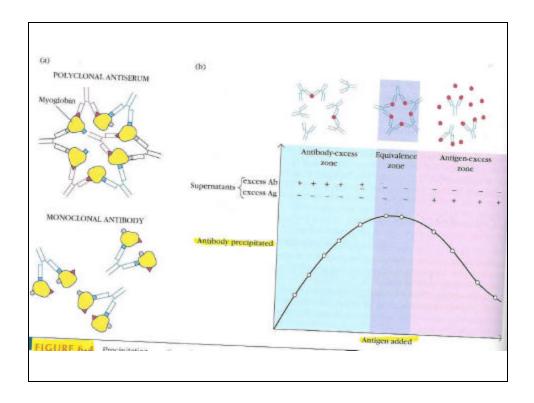
- the Ag must be either bivalent or polyvalent; that is, it must have at least two copies of the same epitope, or have different epitopes that react with different antibodies present in polyclonal antisera

A. Precipitation Reactions in Fluids

- excess of either Ab or Ag interferes with maximal precipitation, which occurs in the equivalence zone, when the ratio of Ab to Ag is optimal

- in the region of Ab excess, unreacted Ab is found in the supernatant along with small, soluble complexes consisting of multiple molecules of Ab bound to a single molecule of Ag

- in the region of Ag excess, unreacted Ag can be detected and small, soluble complexes are again observed, this time consisting of 1 or 2 molecules of Ag bound to a single molecule of Ab



B. Precipitation Reactions in Gels

immunodiffusion reactions can be used

to determine relative concentrations of Abs' or Ags', to compare Ags', or to determine

the relative purity of an Ag preparation

1. Radial Immunodiffusion

- Ag sample is placed in a well and allowed to diffuse into agar containing a suitable dilution of an antiserum

- area of precipitin ring formed is proportional to the concentration of Ag

2. Double Immunodiffusion

- qualitative tool for determining the relationship between antigens and the number of different Ag-Ab systems present

- identity: occurs when two antigens share identical epitopes

- nonidentity: occurs when two antigens are unrelated (share no common epitopes)

- partial identity: occurs when two antigens share some epitopes but one or the other has a unique epitope

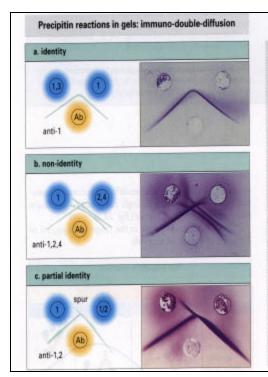
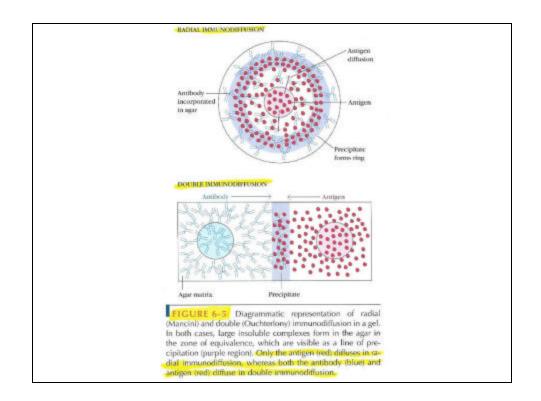


Fig. 28.2 In immuno-double-diffusion, agar gels are pound onto slides and allowed to set; wells are then punched in the geland the test solutions of antigen (Ag) and antibody (Ab) are added. The solutions diffuse out and where Ag and Ab meet they bind to each other, cross-link and precipitate leaving a line of precipitation. The precipitin bands can be visualized by washing the gel to remove soluble proteins and then staining the precipitin arcs with a protein stain such as Coomassie blue. This technique may be used to determine the relationship between antigene (blue) and a particular test antibody (yellow). Three basic patterns appear. The numbers in the blue wells refer to the epitopes present on the test antigen. In reaction (a) the precipitin arcs formed between the antibody and the two test antigens fuse. indicating that the antibody is precipitating identical epitopes in each preparation (epitope 1). This does not mean that the antigers are necessarily identical; they are only identical in as far as the antibody cannot distinguish a difference. In reaction (b) the antibody preparation distinguishes the three different antigens. which form independent precipitin arcs. In reaction (c) the antigens share epitope 1 but one antigen also has epitope 2. This is the same situation as in (a), but in this case the antibody can distinguish them, by virtue of being able to react against both epitopes. A line of identity forms with anti-epitope 1, with the addition of a 'spur' where the anti-epitope 2 has reacted with the second epitope, thus indicating partial rather than total identity between the antigen preparations.



3. Immunoelectrophoresis
- combines separation by electrophoresis with identification by double immunodiffusion
- useful in determining whether a patient produces abnormally low amounts of one or more isotypes, characteristic of certain immunodeficiency diseases
IV. Agglutination Reactions
- interaction between Ab and a particulate Ag results in clumping called agglutination
- Ab excess can inhibit agglutination reactions, which is called prozone effect - at high [Ab], the number of Ab binding sites ma greatly exceed the number of epitopes, which results in antibodies being able to bind antigen only univalently instead of multivalently

- Ab's that bind univalently cannot crosslink one Ag to another

A. Hemagglutination

- agglutination reactions that are performed on RBC's

- RBC's mixed with antisera to the A or B blood-group antigens on a slide

- if antigen is present on the cells, they agglutinate, forming a visible clump on slide

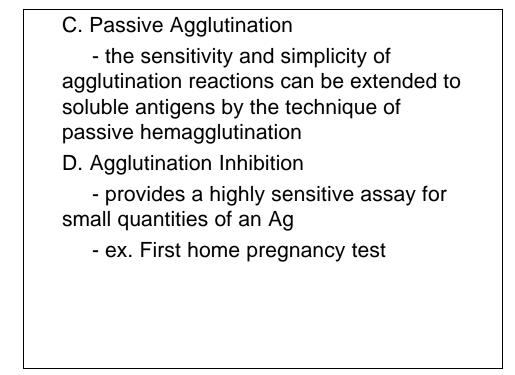
B. Bacterial Agglutination

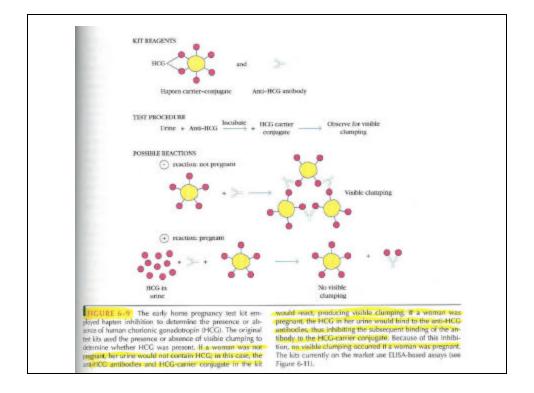
- bacterial infection often elicits the production of serum antibodies specific for surface antigens on the bacterial cells

- presence of such antibodies can be detected by bacterial agglutination reactions

- serum antibody titer of a patient is defined as the reciprocal of the greatest serum dilution that elicits a positive agglutination reaction

- agglutination titer of an antiserum can be used to diagnose a bacterial infection





V. Radioimmunoassay

- one of the most sensitive techniques for detecting antigen or antibody

- principle involves competitive binding of radiolabeled Ag and unlabeled Ag to a high affinity Ab

- ratio of Ab to radioactive Ag is chosen such that the number of epitopes presented by the labeled Ag always exceeds the total number of Ab binding sites

- this insures that any unlabeled Ag added to the sample mixture will compete with radiolabeled Ag for the limited supply of Ab

VI. Enzyme-Linked Immunosorbent Assay

- also known as ELISA

- an enzyme conjugated with an Ab reacts with a colorless substrate to generate a colored reaction product

- substrate known as chromogenic substrate

A. Indirect ELISA

- Ab can be detected or quantitatively determined

- method of choice to detect the presence of serum antibodies against HIV

B. Sandwich Assay

- Ab (rather than Ag) is immobilized on a microtiter well

- sample containing Ag is added and allowed to react with immobilized Ab

- After well is washed, a second enzyme-linked Ab specific for a different epitope on the Ag is added and allowed to react with the bound Ag

- after any free 2nd Ab is removed by washing, substrate is added, and the colored reaction product is measured

C. Competitive ELISA

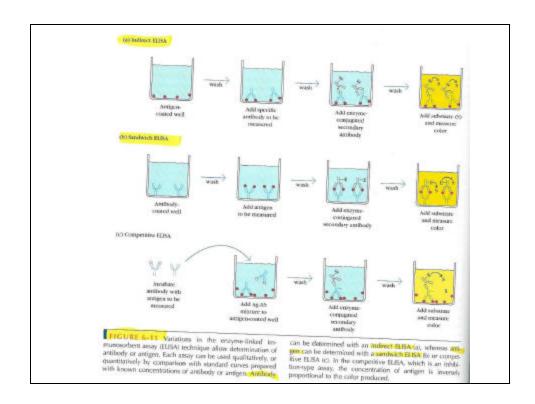
- Ab is first incubated in solution with a sample containing Ag

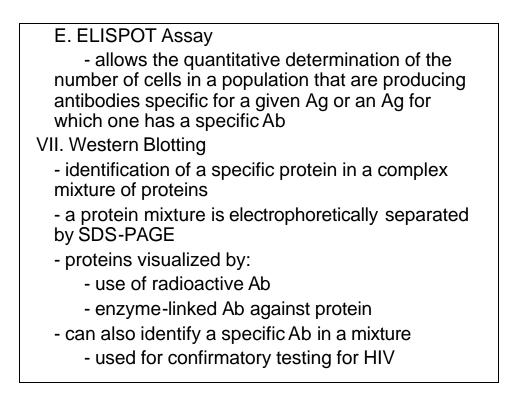
- Ag-Ab mixture is then added to an Agcoated microtiter well

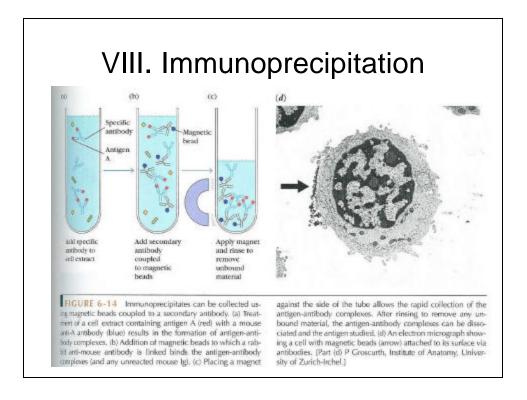
- addition of an enzyme-conjugated secondary Ab specific for the isotype of the primary Ab can be used to determine the amount of primary Ab bound to the well as in an indirect ELISA

D. Chemiluminescence

- a luxogenic (light-generating) substrate takes the place of the chromogenic substrate in conventional ELISA reactions







IX. Immunofluorescence

- most commonly used fluorescent dyes are fluorescein and rhodamine, but other highly fluorescent substances (phycoerythrin and phycobiliproteins) have also come into use

- fluorescein: absorbs blue light and emits an intense yellow-green fluorescence

- rhodamine: absorbs yellow-green range and emits a deep red fluorescence

- phycoerythrin: efficient absorbers of light and brilliant emitters of red fluorescence X. Flow Cytometry and Fluorescence

- FACS (fluorescence-activated cell sorter) was designed to automate the analysis and separation of cells stained with fluorescent Ab

- FACS used to determine the kind and number of WBC's in each population in patients' blood samples

- FACS is one of the essential tools for the detection and classification of leukemias

XI. Immunoelectron Microscopy

- used to visualize specific intracellular tissue components by using the specificity of antibodies